

REDESCRIPTIONS OF *TETRALIA CAVIMANA*
HELLER, 1861 AND *TRAPEZIA CYMODOCE*
(HERBST, 1799) FIRST STAGE ZOEAS WITH
IMPLICATIONS FOR CLASSIFICATION WITHIN
THE SUPERFAMILY XANTHOIDEA
(CRUSTACEA: BRACHYURA)

Paul F. Clark and Bella S. Galil

Abstract.—The first stage zoea of *Tetralia glaberrima* (Herbst, 1790) (now *T. cavimana* Heller, 1861) and *Trapezia cymodoce* (Herbst, 1799) of Gurney (1938) are redescribed and compared both with the original description and with the description of *Tetralia glaberrima* by Al-Kholy (1963). Differences between the two larvae are tabulated and larval characters that appear to support the separation of the families Platyxanthidae and Trapeziidae, as proposed in a classification of adult xanthoids by Guinot (1977 and 1978), are identified.

Guinot (1978) proposed a new classification of adult Brachyura based primarily on the position of female and male genital openings. Eight families were recognized within the superfamily Xanthoidea, and she supported Ortmann (1897) in giving the rank of family to the Trapeziinae as defined by Miers (1886). But for a few minor changes, Guinot indicated that the family contained all the genera listed by Balss (1957) in his subfamily Trapeziinae. Guinot did not use any larval characters to corroborate her classification.

Rice (1980) and Martin (1984) related larval groupings, based on chaetotaxy, to various classifications of adult xanthoids. Both found that many existing descriptions of xanthoid larvae were inadequate or unreliable, and each attempted to use larval evidence to resolve incongruences between different schemes of adult classification. Rice found that his larval groupings did not correspond to the simple divisions of the Xanthidae by Balss (1957), while Martin (1984) endorsed the scheme proposed by Glaessner (1969) because it was based on fossil and recent forms. But later, Martin et al. (1985)

stated that evidence appeared partly to support Balss, as their group I corresponded to the subfamily Xanthinae. Rice (1980) and Martin et al. (1985) agreed that the genus *Homalaspis* warranted separation from the remainder of the Xanthinae of Balss and thereby corroborated the Platyxanthidae of Guinot. Rice therefore tended to support the more complex divisions suggested by Guinot.

The larval descriptions of *Tetralia glaberrima* (Herbst, 1790) (now *T. cavimana* Heller, 1861; see synonymy of Galil, 1988) and *Trapezia cymodoce* (Herbst, 1799) by Gurney (1938) and of *Tetralia glaberrima* by Al-Kholy (1963) are incomplete. Therefore, the aim of this paper is to redescribe these larvae and use this information to re-examine the classification of the xanthoids.

Materials and Methods

Material collected and hatched at the Biological Station, Ghardaqa, Egypt, by Gurney (1938) was fixed originally in formalin and recently transferred to 70% ethanol. The female and the first zoea of *Trapezia cy-*

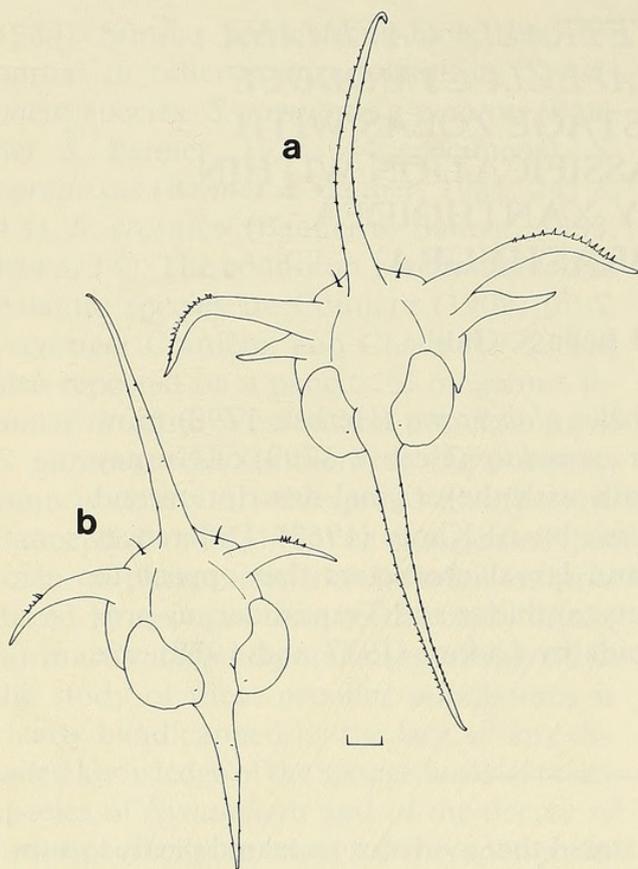


Fig. 1. Carapace of a, *Tetralia cavimana*; b, *Trapezia cymodoce*. Scale bar = 0.1 mm.

modoce (Herbst) are stored in the British Museum (Natural History), registration number 1986:915, and those of *Tetralia cavimana* Heller, were registered as 1986:53. Dissected appendages were mounted in polyvinyl lactophenol and examined using an Olympus BH-2 microscope with Nomarski interference contrast. Drawings were made with the aid of a camera lucida. When comparing the first stage zoea of the two species, the setal arrangement on many appendages was similar and it was only necessary to figure fully the chaetotaxy of one species, *Tetralia cavimana*, and illustrate the differences in *Trapezia cymodoce*.

Tetralia cavimana Heller, 1861

Figs. 1a, 2a–d, 3a, b, 4c, d

Tetralia glaberrima (Herbst, 1790).—Gurney, 1938:77, pl. III, figs. 29–33.—Al-Kholy, 1963:138, pl. I, figs. 1–7.—Williamson, 1970:37.

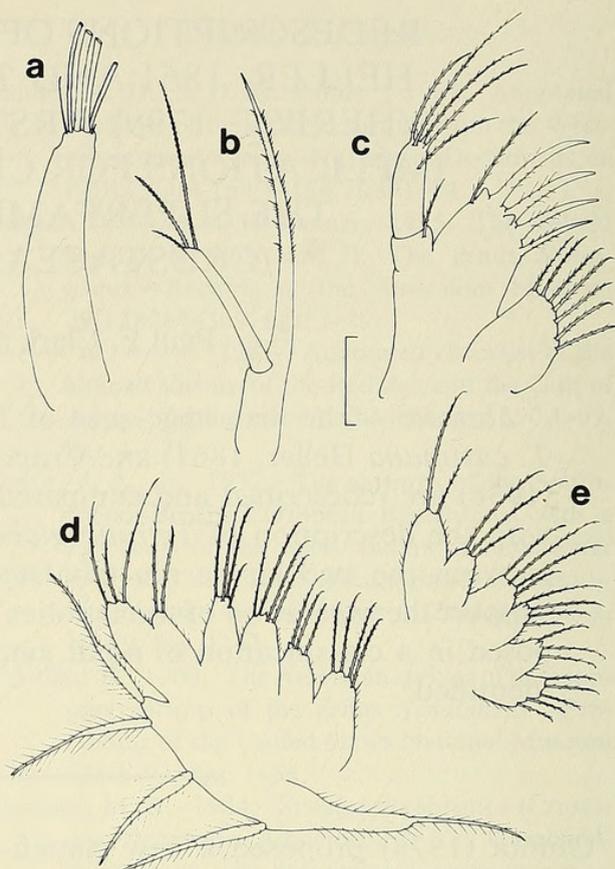


Fig. 2. *Tetralia cavimana*: a, Antennule; b, Antenna; c, Maxillule; d, Maxilla. *Trapezia cymodoce*: e, Setae of maxilla coxal, basal and endopod lobes. Scale bar = 0.1 mm.

non *Tetralia glaberrima* (Herbst, 1790).—Al-Kholy 1963:139–140, pl. I, figs. 8–21, pl. II, figs. 22–33.

First zoea.—Carapace (Fig. 1a): Long dorsal and rostral spines spinulate; 2 pairs of lateral spines, dorsal pair j-shaped and spinulate on dorsal margin, ventral pair smaller than dorsal pair, unarmed; one pair of posterodorsal setae; ventral margin of carapace with minute denticles, marginal setae absent; one pair of posterodorsal carapace setae; eyes sessile with small setule on each eye.

Antennule (Fig. 2a): Endopod absent; exopod unsegmented with 4 terminal aesthethascs, 1 terminal seta and 1 minute terminal spine.

Antenna (Fig. 2b): Spinous process distally spinulate; endopod absent; exopod with unequal terminal setae.

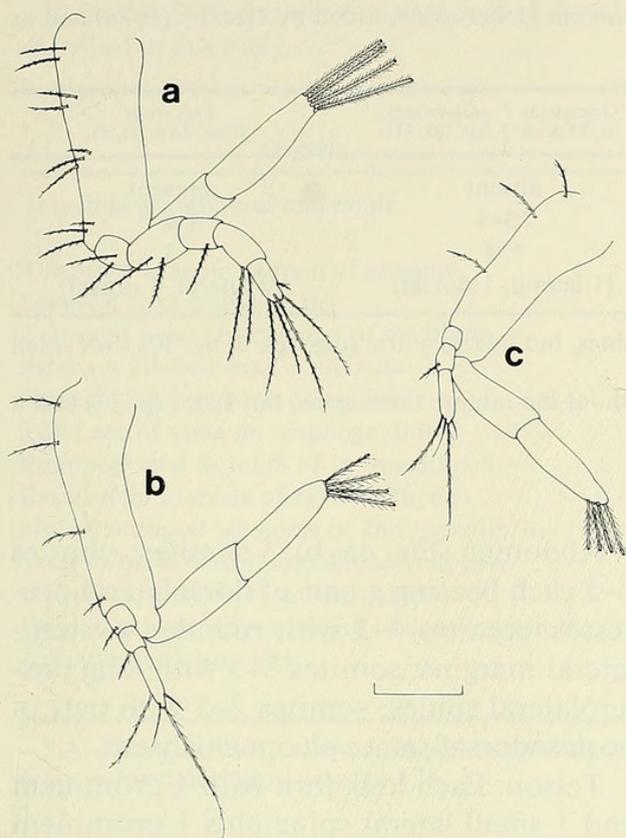


Fig. 3. *Tetrulia cavimana*: a, First maxilliped; b, Second maxilliped. *Trapezia cymodoce*: c, Second maxilliped. Scale bar = 0.1 mm.

Mandible: Endopod (palp) absent.

Maxillule (Fig. 2c): Coxal endite with 7 setae; basal endite with 5 processes on inner margin plus 2 minute teeth, single seta absent from outer margin; endopod 2-segmented, proximal segment with 1 seta, distal segment with 1 subterminal and 4 terminal setae.

Maxilla (Fig. 2d): Coxal endite bilobed with 4+3(+1 minute tooth) setae; basal endite bilobed with 4(+1 minute tooth)+4 setae; endopod bilobed with 2+3 setae; scaphognathite (exopod) with 4 marginal setae plus 1 long stout posterior seta.

First maxilliped (Fig. 3a): Basis with 10 setae arranged 2,2,3,3; endopod 5-segmented with 2,2,1,2,5 setae respectively; exopod 2-segmented, distal segment with 4 terminal natatory setae.

Second maxilliped (Fig. 3b): Basis with 4 setae; endopod 3-segmented with 1,1,4 se-

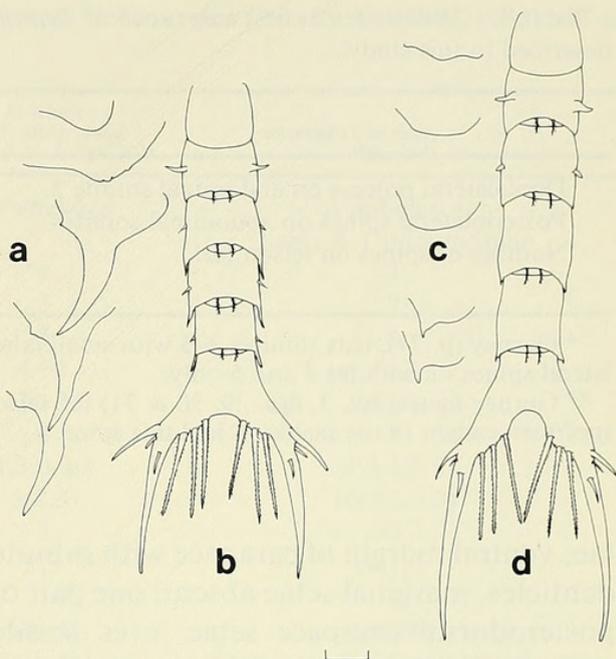


Fig. 4. *Trapezia cymodoce*: a, Dorsolateral spines of somites 1-5; b, Abdomen from dorsal aspect. *Tetrulia cavimana*: c, Dorsolateral spines of somites 1-5; d, Abdomen from dorsal aspect. Scale bar = 0.1 mm.

tae respectively; exopod 2-segmented, distal segment with 4 terminal natatory setae.

Third maxilliped: Undeveloped.

Pereiopods: Undeveloped.

Abdomen (Fig. 4c, d): 5 somites; somites 2 and 3 each bearing 1 pair of dorsolateral processes; somites 1-3 with rounded posterolateral margins; somites 4 and 5 with small posterolateral spines; somites 2-5 with pair of posterodorsal setae; pleopods absent.

Telson (Fig. 4d): Each long fork with 1 prominent and 1 small lateral spine plus 1 prominent dorsal spine; posterior margin with 3 pairs of setae.

Trapezia cymodoce (Herbst, 1799)

Figs. 1b, 2e, 3c, 4a, b

Trapezia cymodoce (Herbst, 1799).—Gurney, 1938:76, pl. II, figs. 23-28.

First zoea.—Carapace (Fig. 1b): long dorsal and shorter rostral spine lightly spinulate; single pair of curved, lightly spinulate lateral spines; one pair of posterodorsal se-

Table 1.—Differences in first stage zoea of *Tetralia cavimana* Heller as described by Gurney (1938) and as described in this study.

Character	Gurney (as <i>T. glaberrima</i>) (p. 77 & pl. 3, figs. 30, 31)	This study (Figs. 3d & 4c, d)
Dorsolateral process on abdominal somite 3	absent	present
Posteriolateral spines on abdominal somites	*3–5	4 & 5
Number of spines on telson fork	**2 (1 lateral, 1 dorsal)	3 (2 lateral, 1 dorsal)

* Gurney (p. 77), lists somites 3–5 with small lateral spines, but figures a first stage (pl. 3, fig. 30) with small lateral spines on somites 4 and 5 only.

** Gurney figures (pl. 3, figs. 29, 30 & 31) the telson without the minute third spine, but states (p. 77) that a specimen caught in the plankton had this spine.

tae; ventral margin of carapace with minute denticles, marginal setae absent; one pair of posterodorsal carapace setae; eyes sessile with small setule on each eye.

Antennule: Endopod absent; exopod unsegmented with 4 terminal esthetascs, 1 terminal seta and 1 minute terminal spine.

Antenna: Spinous process distally spinulate; endopod absent; exopod with unequal terminal setae.

Mandible: Endopod (palp) absent.

Maxillule: Coxal endite with 7 setae; basal endite with 5 processes on inner margin plus 2 minute teeth, single seta absent from outer margin; endopod 2-segmented, proximal segment with 1 seta, distal segment with 1 subterminal and 4 terminal setae.

Maxilla (Fig. 2e): Coxal endite bilobed with 5+3(+1 minute tooth) setae; basal endite bilobed with 4(+1 minute tooth)+4 setae; endopod bilobed with 3+2 setae; scaphognathite (exopod) with 4 marginal setae plus 1 long stout posterior seta.

First maxilliped: Basis with 10 setae arranged 2,2,3,3; endopod 5-segmented with 2,2,1,2,5 setae respectively; exopod 2-segmented, distal segment with 4 terminal natatory setae.

Second maxilliped (Fig. 3c): Basis with 3 setae; endopod 3-segmented with 0,1,4 setae respectively; exopod 2-segmented, distal segment with 4 terminal natatory setae.

Third maxilliped: Undeveloped.

Pereiopods: Undeveloped.

Abdomen (Fig. 4a, b): 5 somites; somites 2–5 each bearing a pair of dorsolateral processes; somites 1–3 with rounded posterolateral margins; somites 3–5 with long posterolateral spines; somites 2–5 with pair of posterodorsal setae; pleopods absent.

Telson: Each long fork with 1 prominent and 1 small lateral spine plus 1 prominent dorsal spine; posterior margin with 3 pairs of setae.

Discussion

The differences between the descriptions by Gurney (1938) and Al-Kholy (1963) of *Tetralia cavimana* Heller (as *T. glaberrima* Herbst) and the present study are listed in Tables 1 and 2. In his description of *Trapezia cymodoce* Gurney (1938) figured only abdominal somites 2 and 3 with dorsolateral processes, but on re-examination of this material they were found additionally on somites 4 and 5 (Fig. 4b). Although the first stage zoeas of *Trapezia guttata* Rüppell, 1830 and *Trapezia maculata* (MacLeay, 1838) have been described by Gurney (1938) and Al-Kholy (1963) respectively, these descriptions are inadequate for comparison. Differences in appendage chaetotaxy between *Trapezia cymodoce* (Herbst) and *Tetralia cavimana* Heller first stage zoeas are listed in Table 3.

Rice (1980), from larval descriptions, divided the Xanthidae into 4 groups. He com-

Table 2.—Differences in first stage zoea of *Tetralia cavimana* Heller as described by Al-Kholy (1963) and as described in this study.

Character	Al-Kholy (as <i>T. glaberrima</i>) (p. 138 & pl. 1, figs. 2-7)	This study (figs. 2-4)
Terminal armature of antennule	2 esthetascs	4 esthetascs 1 seta & 1 minute spine
Terminal setae on exopod of antenna	*4	3
Setae on coxa of maxillule	5	7
Terminal setae on endopod of maxillule	3	4
Setae on bilobed coxa of maxilla	4+5	4+3
Setae on bilobed basis of maxilla	2+3	4+4
Total no. of setae on scaphognathite	4	5
Endopod setal formula of 1st maxilliped	1,3,0,0,3	2,2,1,2,5
Basial setal formula of 1st maxilliped	5(2,3)	10(2,2,3,3)
No. of endopod segments of 2nd maxilliped	2	3
Setal formula of 2nd maxilliped endopod	2,3	1,1,4
No. of basal setae of 2nd maxilliped	1	4
Dorsolateral processes on 3rd abdominal somite	absent	present
Armature of telson fork	1 prominent lateral spine	1 prominent & 1 small lateral spine + 1 prominent dorsal spine

* Al-Kholy figures 3 terminal setae on antenna exopod (pl. 1, fig. 2), but scores an exopod with 4 terminal setae in his description on page 138.

pared these groupings with several adult classifications, and concluded that they gave some support to the divisions proposed by Guinot (1978). Martin (1984) recognized six groups (I–VI) within the Xanthidae, based on zoeal characters, but gave his groupings no formal taxonomic status. He adopted the classification of xanthoids proposed by Glaessner (1969), because his larval groups did not correspond to the eight families of Guinot.

Rice (1980) and Martin (1984) agree on the suite of characters that defines their xanthoid group III. The genera that they assigned to their respective groups III differs. *Eriphia*, *Homalaspis*, *Ozius* and *Tetralia* form the group III of Rice. The group III of Martin contains *Baptozius*, *Carpilius*, *Epixanthus*, *Paramedaeus*, *Pilumnoides*, *Platyxanthus* and *Trapezia* in addition to those of Rice. Both include ASM 26 in group III. However, the present redescription of *Trapezia cymodoce* (Herbst) and *Tetralia cavimana* Heller first stage zoeas indicates that the group III of both workers can be

divided into two subgroups A and B, the characters of which are defined in Table 4.

Group A includes the first stages zoeas of *Tetralia cavimana* Heller and *Trapezia cymodoce* (Herbst) and corresponds to the Trapeziidae of Guinot (1978). Group B comprises her Platyxanthidae and contains the zoeas of *Homalaspis plana* (A. Milne Edwards) (Fagetti 1970), *Platyxanthus*

Table 3.—Differences in chaetotaxy of appendages between first zoea stages of *Trapezia cymodoce* (Herbst) and *Tetralia cavimana* Heller.

Character	<i>Trapezia cymodoce</i>	<i>Tetralia cavimana</i>
No. of lateral carapace spines	1	2
Setation of maxilla endopod	3+2	2+3
Setation of maxilla coxa	5+3	3+4
Setal formula of endopod of 2nd maxilliped	0,1,4	1,1,4
Basial setae of 2nd maxilliped	3	4
Abdominal somites with dorso-lateral processes	2-5	2 & 3
Abdominal somites with dorso-lateral spines	3-5 (long)	4 & 5 (short)

Table 4.—Proposed characters that subdivide the xanthid group III of Martin (1984) and Rice (1980).

Character	Group A	Group B
Setation of distal endopod segment of maxillule	1 subterminal + 4 terminal	2 subterminal + 4 terminal
Setation of maxilla endopod	subterminal setae absent (2 or 3 terminal setae only)	subterminal setae present (2 subterminal + 3 terminal setae)
Setation of basal endopod segment of 1st maxilliped	2	*3
Setation of distal endopod segment of 2nd maxilliped	4	**6

* Lumare & Gozzo (1972) figure the zoeal stages of *Eriphia verrucosa* as variously having 2 or 3 setae on this segment. Examination of *E. verrucosa* 1st stage zoeas from Ischia, Italy (plankton caught material by Galil) revealed 3 setae in this position. Hashmi (1970) described and figured the 1st zoeal stage of *Eriphia laevimana smithii* (MacLeay) with 2 setae, but re-examination of his material, BM(NH) registration number 1986:908, also revealed 3 setae. Wear (1968) illustrated the 1st maxilliped of *Ozius truncatus* H. Milne Edwards stage I zoea with only 1 seta on the basal endopod segment. Other zoeas in this group have 3 setae.

** The following 1st stage zoea all have 6 setae present on the distal segment of second maxilliped endopod; *Baptozius vinosus* (H. Milne Edwards), *Eriphia laevimana smithii* MacLeay, *Eriphia verrucosa* (Forskål), *Homalaspis plana* (A. Milne Edwards), *Monodeus couchii* (Couch) & *Platyxanthus patagonicus* A. Milne Edwards.

crenulatus (A. Milne Edwards) (Menu-Marque 1970) and *P. patagonicus* A. Milne Edwards (Iorio & Boschi 1986). Guinot (1978) also lists the genera *Homalaspis* and *Platyxanthus* in her Platyxanthidae. Other larvae that fit in group B include the menippids *Baptozius vinosus* (H. Milne Edwards) (Saba et al. 1978a), *Epixanthus dentatus* (White) (Saba et al. 1978b), *Eriphia laevimana smithii* MacLeay (Hashmi 1970), *E. spinifrons* (Herbst) (Bourdillon-Casanova 1960; Hyman 1925; Paolucci 1910), *E. verrucosa* (Forskål) (Lumare and Gozzo 1972), *Ozius rugulosus* Stimpson (Kakati and Nayak 1977), *O. truncatus* H. Milne Edwards (Wear 1968) and the pilumnid *Pilumnoides perlatus* (Poëppig) (Fagetti & Campodonico 1973). None of these are listed in Guinot's classification, but on the basis of larval characters these genera appear to have affinities with the Platyxanthidae. The xanthids *Monodaeus couchii* (Couch) (Ingle 1983) and *Paramedaeus noelensis* (Ward) (Suzuki 1979) are also grouped in the Platyxanthidae. In her classification of adult Xanthidae, Guinot (1978) placed *Monodaeus* and *Paramedaeus* in the sub-family Euxanthinae Alcock, 1898. Apart from this anomaly, the

larval evidence presented here appears to correspond to the classification of adult Platyxanthidae and Trapeziidae as proposed by Guinot (1978).

The status of the third stage zoea of "ASM 26 (Menippinae or Trapeziinae)," as described by Rice and Williamson (1977:52–54, fig. 27) remains uncertain; it does not have a menippid type antenna, maxillule or maxilla endopod. Martin (1984) believed that "ASM 26" was more likely to be in the Trapeziinae, but from evidence presented here it does not appear to share the characters defining group A (see Table 4).

Carpilius was placed by Martin et al. (1985) in xanthid group III. Laughlin et al. (1983) described the zoeal stages of the coral crab *Carpilius corallinus* (Herbst) and noted a number of diagnostic characters: 5 zoeal stages, the separation of the 6th abdominal somite from the telson in stage II zoea, appearance of pleopod buds in stage II, the large size of the zoea, the increased numbers of natatory setae in the maxillipeds in later stages (i.e., zoea IV with 15–16 and zoea V with 20–22 natatory setae), and the large size of the mandibles. In combination, these larval characters are interpreted by Laugh-

lin et al. (1983) as evidence supporting the establishment of the family Carpiliidae by Guinot (1978).

Careful re-examination of other xanthoidan larval descriptions may further substantiate the classification proposed by Guinot (1978).

Literature Cited

- Al-Kholy, A. A. 1963. The zoeal stages of *Tetralia glaberrima* (Herbst), from the Red Sea.—Publications of the Marine Biological Station, Ghardaq, Red Sea, Cairo No. 12:137–144.
- Balss, H. 1957. Decapoda VIII Systematik in H. G. Bronn, Klassen und Ordnungen des Tierreichs Band 5, Abteilung 1, 7(12):1505–1672.
- Bourdillon-Casanova, L. 1960. Le méroplankton du Golfe de Marseille: les larves de Crustacés Décapodes.—Recueil des Travaux de la Station d'Endoume, Marseille 30:1–286.
- Fagetti, E. 1970. Desarrollo larval en el laboratorio de *Homalaspis plana* (Milne Edwards) (Crustacea, Brachyura, fam. Xanthidae).—Revista de Biología Marina, Valparaiso 14:29–49.
- , & I. Campodonico. 1973. Larval development of *Pilumnoides perlatus* (Brachyura: Xanthidae) under laboratory conditions. Marine Biology. First International Interdisciplinary Conference on Marine Biology, Washington 18: 129–139.
- Galil, B. S. 1988. Trapeziidae (Decapoda: Brachyura: Xanthoidea) of the Red Sea, Israel Journal of Zoology, Jerusalem 34(314):159–182.
- Glaessner, M. F. 1969. Decapoda. Pp. 399–533 in R. C. Moore, ed., Treatise on invertebrate paleontology Part R, Arthropod 2. Geological Society of America and University of Kansas, Connecticut & New York.
- Guinot, D. 1977. Propositions pour une nouvelle classification des Crustacés Décapodes Brachyours.—Compte Rendu des Séances de la Société de Biologie, Paris sér. D, 285:1049–1052.
- . 1978. Principes d'une classification évolutive des Crustacés Décapodes Brachyours.—Bulletin Biologique de la France et de la Belgique 112(3):211–292.
- Gurney, R. 1938. Notes on some decapod Crustacea from the Red Sea. VI–VIII.—Proceedings of the Zoological Society of London Ser. B. 108:73–84.
- Hashmi, S. S. 1970. Study on the larvae of the family Xanthidae (*Heteropanope*, *Eurycarcinus*, and *Eriphia*) hatched in the laboratory. Brachyura: Decapoda.—Agriculture Pakistan, Karachi 21(4):457–478.
- Hyman, O. W. 1925. Studies on the crabs of the family Xanthidae.—Proceedings of the United States National Museum 67:1–22.
- Ingle, R. W. 1983. A comparative study of the larval development of *Monodaeus couchi* (Couch), *Xantho incisus* Leach and *Pilumnus hirtellus* (Linnaeus) (Crustacea: Brachyura: Xanthidae).—Journal of Natural History, London 17: 951–978.
- Iorio, I. M., & E. E. Boschi. 1986. Studies on the larval stages of the crab *Platyxanthus patagonicus*, from laboratory rearing and plankton samples.—Journal of Agriculture in the Tropics 1(1): 7–24.
- Kakati, V. S., & V. N. Nayak. 1977. Larval development of the xanthid crab, *Ozius rugulosus* Stimpson (Decapod, Brachyura) under laboratory conditions.—Indian Journal of Marine Sciences, New Delhi 6:26–30.
- Laughlin, R. A., P. J. Rodriguez, & J. A. Marval. 1983. Zoeal stages of the coral crab *Carpilius corallinus* (Herbst) (Decapoda, Xanthidae) reared in the laboratory.—Crustaceana 44(2):169–186.
- Lumare, F., & S. Gozzo. 1972. Sviluppo larvale de Crostaceo Xanthidae *Eriphia verrucosa* (Forskål, 1775) in condizioni di laboratorio.—Bollettino di Pesca, di Piscicoltura ed Idrobiologica, Roma 27(1):185–209.
- Martin, J. W. 1984. Notes and bibliography on the larvae of xanthid crabs, with a key to the known xanthid zoeas of the western Atlantic and Gulf of Mexico.—Bulletin of Marine Science 34(2): 220–239.
- , F. M. Truesdale, & D. L. Felder. 1985. Larval development of *Panopeus bermudensis* Benedict and Rathbun, 1891 (Brachyura, Xanthidae) with notes on zoeal characters in xanthid crabs.—Journal of Crustacean Biology 5(1):84–105.
- Menu-Marque, S. A. 1970. Desarrollo larval de cangrejo *Platyxanthus crenulatus* (A. Milne Edwards, 1879) en el laboratorio (Decapoda, Brachyura, Xanthidae).—Physis, Buenos Aires 29(79):477–494.
- Miers, E. J. 1886. Report on the Brachyura collected by H.M.S. Challenger during the years 1873–1876. Report on the Scientific Results of the voyage of H.M.S. Challenger during the years 1873–76, Zoology 17:xli + 362 pp.
- Ortmann, A. E. 1897. Die geographische Verbreitung de Decapoden-Familie Trapeziidae.—Zoologische Jahrbucher. Abteilung fur Systematik 10: 201–216.
- Paolucci, C. 1910. La zoea di *Eriphia spinifrons*.—Rivista Mensile Pesca, Pavia 5:33–40.
- Rice, A. L. 1980. Crab zoeal morphology and its bearing on the classification of the Brachyura.—Transactions of the Zoological Society of London 35:271–424.
- , & Williamson, D. I. 1977. Planktonic stages

- of Crustacea Malacostraca from Atlantic sea-mounts.—“Meteor” Forschungsergebnisse, Gebriider Borntraeger, Berlin-Stuttgart Reihe D, Biologie, No. 26:28–64.
- Saba, M., M. Takeda, & Y. Nakasone. 1978a. Larval development of *Baptozius vinosus* (H. Milne Edwards).—Proceedings of the Japanese Society of Systematic Zoology, Tokyo 14:25–38.
- , ———, & ———. 1978b. Larval development of *Epixanthus dentatus* (White) (Brachyura, Xanthidae).—Bulletin of the National Science Museum (Zoology), Tokyo 4:151–161.
- Suzuki, H. 1979. Studies on the zoea of the two xanthid crabs, *Paramedaeus truncatus* (de Haan) (Crustacea, Brachyura, Xanthidae).—Proceedings of the Japanese Society of Systematic Zoology 16:35–52.
- Wear, R. G. 1968. Life history studies on New Zealand Brachyura 2. Family Xanthidae. Larvae of *Heterozius rotundifrons* A. Milne Edwards, 1867, *Ozius truncatus* H. Milne Edwards, 1834, and *Heteropanope (Pilumnopeus) serratifrons* (Kinahan, 1856).—New Zealand Journal of Marine and Freshwater Research 2:698–707.
- Williamson, D. I. 1970. On a collection of planktonic Decapoda and Stomatopoda (Crustacea) from the east coast of the Sinai Peninsula, northern Red Sea.—Bulletin of the Sea Fisheries Research Station, Israel No. 56:3–48.
- (PFC) Department of Zoology, British Museum (Natural History), Cromwell Road, London SW7 5BD, England; (BSG) Department of Zoology, George S. Wise Faculty of Life Sciences, Tel Aviv University, Israel.



Clark, Paul F and Galil, Bella. 1988. "Redescriptions Of Tetralia cavimana Heller, 1861 And Trapezia cymodoce Within The Superfamily Xanthoidea (Crustacea, Brachyura)." *Proceedings of the Biological Society of Washington* 101, 853–860.

View This Item Online: <https://www.biodiversitylibrary.org/item/107746>

Permalink: <https://www.biodiversitylibrary.org/partpdf/46453>

Holding Institution

Smithsonian Libraries and Archives

Sponsored by

Biodiversity Heritage Library

Copyright & Reuse

Copyright Status: In copyright. Digitized with the permission of the rights holder.

Rights Holder: Biological Society of Washington

License: <http://creativecommons.org/licenses/by-nc-sa/3.0/>

Rights: <https://biodiversitylibrary.org/permissions>

This document was created from content at the **Biodiversity Heritage Library**, the world's largest open access digital library for biodiversity literature and archives. Visit BHL at <https://www.biodiversitylibrary.org>.