Evolution of the Toxoglossa Venom Apparatus as Inferred by Molecular Phylogeny of the Terebridae

Mandë Holford,*† Nicolas Puillandre,‡ Yves Terryn,§ Corinne Cruaud,|| Baldomero Olivera,* and Philippe Bouchet‡

*Biology Department, University of Utah; †Chemistry Department, The City University of New York-York College; †Muséum National d'Histoire Naturelle (MNHN), Departement Systematique et Evolution, Unité Mixte de Recherche 7138 Systématique, Adaptation, Evolution, 55 rue Buffon, 75231 Paris Cedex 05, France; NaturalArt, Gent, Belgium; and ||GENOSCOPE, Centre National de Séquençage, Evry, France

Toxoglossate marine gastropods, traditionally assigned to the families Conidae, Terebridae, and Turridae, are one of the most populous animal groups that use venom to capture their prey. These marine animals are generally characterized by a venom apparatus that consists of a muscular venom bulb and a tubular venom gland. The toxoglossan radula, often compared with a hypodermic needle for its use as a conduit to inject toxins into prey, is considered a major anatomical breakthrough that assisted in the successful initial radiation of these animals in the Cretaceous and early Tertiary. The pharmacological success of toxins from cone snails has made this group a star among biochemists and neuroscientists, but very little is known about toxins from the other Toxoglossa, and the phylogeny of these families is largely in doubt. Here we report the first molecular phylogeny for the Terebridae and use the results to infer the evolution of the venom apparatus for this group. Our findings indicate that most of the genera of terebrids are polyphyletic, and one species ("Terebra" (s.l.) jungi) is the sister group to all other terebrids. Molecular analyses combined with mapping of venom apparatus morphology indicate that the Terebridae have lost the venom apparatus at least twice during their evolution. Species in the genera Terebra and Hastula have the typical venom apparatus found in most toxoglossate gastropods, but all other terebrid species do not. For venomous organisms, the dual analysis of molecular phylogeny and toxin function is an instructive combination for unraveling the larger questions of phylogeny and speciation. The results presented here suggest a paradigm shift in the current understanding of terebrid evolution, while presenting a road map for discovering novel terebrid toxins, a largely unexplored resource for biomedical research and potential therapeutic drug development.

Introduction

Marine snails are not what initially come to mind when discussing venomous animals, but the toxoglossan gastropods that use venom to capture their prey are among the most highly populous groups of marine invertebrates. Toxoglossate ("poison tongue") gastropods include cone snails (Conus, within the family Conidae), auger snails or Terebridae, and "Turridae" (a complex of families) (Taylor et al. 1993; Puillandre et al. 2008). A venom apparatus made up of a muscular venom bulb and a tubular venom gland generally characterizes these marine mollusks. Toxins are injected into target animals via hollow disposable radular teeth and act to immobilize prey or defend against predators (Olivera 2002). The toxoglossan radula, often compared with a hypodermic needle, is considered a major anatomical breakthrough that assisted in the successful initial radiation of these animals in the Cretaceous and lower Tertiary (Shimek and Kohn 1981). There are believed to be >10,000 species of toxoglossate mollusks (Bouchet 1990); Toxoglossa are the most diverse and abundant group of predatory snails in species numbers. The pharmacological success of toxins from cone snails has led to their extensive use in characterizing cellular communication in the nervous system, but very little is known about toxins from terebrids and other Toxoglossa, and the phylogeny of these families is largely in doubt.

The Terebridae are a distinctive example of modular anatomical development within marine gastropods, with their foregut anatomy possessing various interchangeable

Key words: Toxoglossa evolution, Terebridae phylogeny, venomous marine snails, peptide toxins, venom apparatus.

E-mail: mholford@york.cuny.edu.

Mol. Biol. Evol. 26(1):15-25. 2009 doi:10.1093/molbev/msn211 Advance Access publication October 6, 2008

parts. All Terebridae are carnivorous and hunt prey; however, a significant number of terebrid species have lost the venom apparatus. Based on key structural differences in their foregut anatomy, Miller (1970, 1971) identified three distinct terebrid feeding types: Type I terebrids, of which there are two groups, IA and IB, have salivary glands, a long eversible labial tube, and a short and slightly retractable buccal tube (fig. 1A). Type I terebrids lack a radular and venom apparatus and therefore may not use toxins to subdue and immobilize their prey. These terebrids feed by grasping the prey with the anterior part of the labial tube and engulfing the prey whole. On the other hand, Type II terebrids exhibit characteristic toxoglossan feeding in that they impale the prey with a hypodermic radular tooth through which they deliver immobilizing toxins expressed in the venom gland, then grasp and engulf their catch with the sphincter of the labial tube. In addition to the hypodermic radular teeth and venom apparatus, these terebrids have salivary glands, an eversible labial tube of moderate length, and a long retractile buccal tube (fig. 1*B*). The final terebrid type, Type III, is similar to Type I in that they lack a venom apparatus but possess an unusual accessory feeding organ, the accessory proboscis structure, that is mostly uncharacteristic in other terebrids (fig. 1C). Type III terebrids use the accessory feeding organ to grasp and ingest tentacles of cirratulid polychates. To understand how the three strategies evolved, it is imperative to establish a molecular phylogeny for the Terebridae.

Similar to the majority of shelled mollusks, the taxonomy of the Terebridae is not very well characterized; their taxonomy is primarily based on shell morphology (Oyama 1961; Bratcher and Cernohorsky 1987; Terryn 2007) and relatively few studies describe their anatomy (Rudman 1969; Miller 1971; Taylor 1990; Taylor et al. 1993). This is surprising, given the relative abundance of terebrids and their global tropical distribution and shallow water

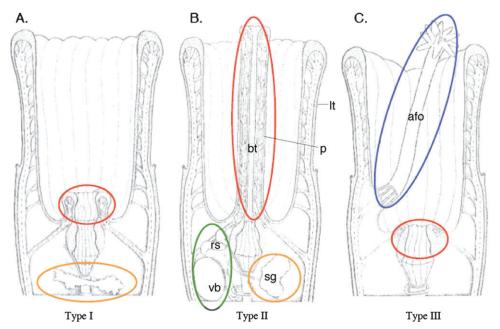


Fig. 1.—Schematic representation of Miller's foregut anatomy of the Terebridae. The three types of foregut anatomy Miller used to define feeding strategies within the Terebridae are depicted (Types I, II, and III). Anatomical features are labeled and highlighted in color: yellow = salivary glands (sg), red = buccal tube (bt), green = venom bulb (vb) and radular sac (rs), and blue = accessory feeding organ (afo). Also labeled are the proboscis (p), found only in Type II terebrids, and the labial tube (lt) (after Miller 1970).

occurrence. The last classification (Bouchet and Rocroi 2005) of the Gastropoda recognizes two subfamilies, Terebridae and Pervicaciinae, within the family Terebridae, which comprises >300 known species (Bratcher and Cernohorsky 1987; Terryn 2007) and may be another 100 unnamed taxa, especially in deeper waters (100–500 m).

We used a three-gene data matrix consisting of mitochondrial 12S and 16S rRNA and cytochrome oxidase sub-unit I (COI) sequences from 67 species to reconstruct the first molecular phylogeny of the Terebridae (fig. 2). We also assessed the evolution of terebrid feeding strategies by mapping the presence or absence of the venom apparatus on the phylogeny. The results presented here suggest a new hypothesis for understanding terebrid evolution: Our findings indicate that the Terebridae have independently lost the venom apparatus twice during their evolution.

The molecular phylogeny of the Terebridae additionally provides a road map for discovering novel terebrid toxins from those species with a venom apparatus. Given the similarities with cone snails, it is expected that the toxins from terebrids may be a rich source of neuroactive peptides to investigate cellular communication in the nervous system, but this is largely unexplored at present. Reconstructing a reliable phylogeny for the Terebridae is the first step toward characterizing their toxins using the emerging biodiversity/exoge-

nome combination strategy that has been developed for *Conus* venom peptides (Olivera and Teichert 2007).

Materials and Methods

Material

The collection details for the nonterebrid samples used in this study are shown in Table 1. All specimens used were collected during several expeditions to the West Pacific since 2004 and specifically fixed for molecular analysis. Living specimens were anesthetized by placing them in a solution with a concentration of 1 volume of MgCl₂ for 13 volume of water, for 1 or 2 h. A piece of tissue (usually foot) was cut and fixed in 95% ethanol. Table 2 lists all terebrid specimens used in this study and the expedition sources. Vouchers are kept in the collection of the Muséum National d'Histoire Naturelle (MNHN). The species included in taxon sampling represent 12 of the 15 identified genera as defined in a recent shell morphology based revision of the Terebridae (Terryn 2007). Outgroups were chosen to form a nonmonophyletic group as recommended by Darlu and Tassy (1993). Representative species of Conidae and Turridae (s.l.) were chosen as closely related outgroups, and one species from the neogastropod family Harpidae was chosen as distant outgroup.

Table 1 Outgroups Used in the Study

MNHN N°	Cruise	Coordinates, Depth	Genus	Species
40568	EBISCO	21°10′S, 158°39′E, 650–723 m	Turridae	Cochlespira sp.
17922	Panglao 05	9°32.5′N, 123°41.8′E, 111–115 m	Conidae	Conus nereis
40569	Santo 06	15°37.3'S, 167°05.8'E, 78–91 m	Harpidae	Harpa sp.
17685	Santo 06	15°33.6'S, 167°16.6'E, 8–9 m	Turridae	Iotyrris cingulifera

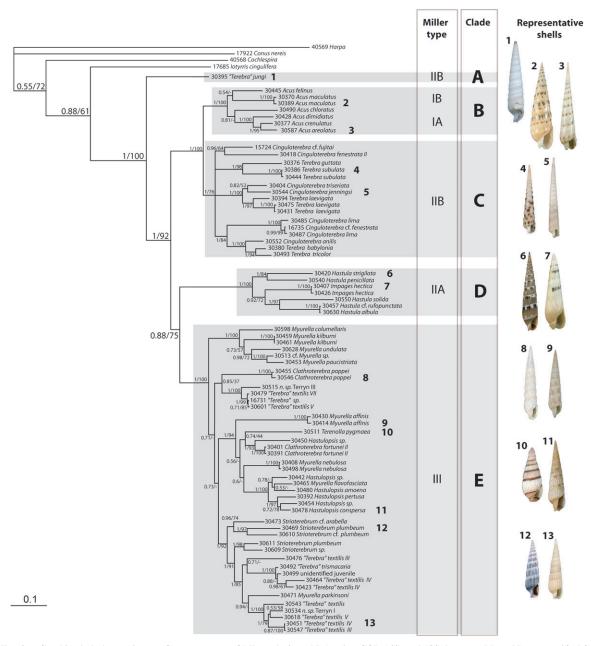


Fig. 2.—Combined phylogenetic tree. Consensus tree of ML analysis and BA using COI, 16S, and 12S data sets. PP and B are specified for each node. Miller types, IA, IB, IIA, IIB, and III, as described in the text are highlighted in the tree. Molecular analyses divide the Terebridae into five distinct clades, Clades A-E, indicated by the shaded gray areas. Representative shells, numbered 1-13, are shown for each clade. For clarity, multiple samples of the same species are shown only when there is a geographic difference, for example, 30370 and 30389 Acus maculatus from Panglao 2004 and Santo 2006 expeditions, respectively.

Sequencing

DNA was extracted from foot or other tissue using Qiagen QIAamp Dneasy Tissue kit. Fragments of mitochondrial genes 12S, 16S, and COI were amplified using universal primers 12S1/12S3 (Simon et al. 1991), 16Sar/16Sbr (Palumbi 1996), and LCO1490/HCO2198 (Folmer et al. 1994), respectively. All polymerase chain reactions (PCRs) were performed in 25 μ l, containing 3 ng of DNA, 1× reaction buffer, 2.5 mM MgCl₂, 0.26 mM deoxynucleoside triphosphate, 0.3 mM each primer, 5% dimethyl sulfoxide, and 1.5 units of Qbiogene Q-Bio Taq. Amplification consisted

of an initial denaturation step at 94 °C for 4 min, followed by 37 cycles of denaturation at 94 °C for 30 s, annealing at 54 °C for 12S gene and 52 °C for 16S, followed by extension at 72 °C for 1 min. The final extension was at 72 °C for 5 min. The amplification of COI genes was performed similarly, except there were two annealing cycles: the first repeated five times at an annealing temperature of 45 °C and the second repeated 30 times at 50 °C as described by Herbert et al. (2003). PCR products were purified using exonuclease I and phosphatase and sequenced using the Applied Biosystem BigDyeTerminator V3.1 kit and the ABI3730XL

Table 2 List of Terebrid Samples Used in Molecular Phylogeny

MNHN N°	Cruise	Coordinates, Depth	Genus	Species (Author)	COI	16S	12S	Miller Type	Clade
30482	BOA1	15°41.6′S, 167°02.1′E, 268–445 m	Cinguloterebra	fujitai (Kuroda and Habe 1952)	X	X			C
30530	EBISCO	20°29′S, 158°42′E, 197–230 m		n. sp. Terryn III	X	X	X		E
30515	EBISCO	20°06′S, 160°23′E, 280–304 m		n. sp. Terryn III	X	X	X		E
30535	Panglao 04	9°33.0′N, 123°46.5′E, 8–14 m	Hastula	lanceata (Linnaeus 1767)	X	X			D
30370	Panglao 04	9°37.4′N, 123°46.9′E, 3–20 m	Acus	maculatus (Linnaeus 1758)	X	X	X	IB	В
30404	Panglao 04	9°35.3′N, 123°52.2′E, 84–87 m	Cinguloterebra	triseriata (JE Gray 1824)	X	X	X		C
30464	Panglao 04	9°35.3′N, 123°52.2′E, 84–87 m	"Terebra" textilis-group	textilis type IV	X	X	X		E
30442	Panglao 04	9°33.5′N, 123°48.6′E, 80–120 m	Hastulopsis	sp.	X	X	X		E
30443	Panglao 04	9°37.4′N, 123°54.5E, 6–8 m	Acus	felinus (Dillwyn 1817)	X	X	X	IB	В
30444	Panglao 04	9°37.4′N, 123°54.5E, 6–8 m	Terebra	subulata (Linnaeus 1767)	X	X	X	IIB	C
30445	Panglao 04	9°37.4′N, 123°54.5E, 6–8 m	Acus	felinus (Dillwyn 1817)	X	X	X	IB	В
30446	Panglao 04	9°37.4′N, 123°54.5′E, 4–5 m	Terenolla	pygmaea (Hinds 1844)	X	X	X	III	E
30448	Panglao 04	9°37.4′N, 123°54.5′E, 4–5 m	Terenolla	pygmaea (Hinds 1844)	X	X	X	III	E
30449	Panglao 04	9°37.4′N, 123°54.5′E, 4–5 m	Terenolla	pygmaea (Hinds 1844)	X	X	X	III	E
30430	Panglao 04	9°37.4′N, 123°54.5E, 6–8 m	Myurella	affinis (JE Gray 1834)	X	X	X	III	E
30452	Panglao 04	9°37.4′N, 123°54.5E, 6–8 m	Myurella	affinis (JE Gray 1834)	X	X	X	III	E
30439	Panglao 04	9°33.4′N, 123°48.4′E, 3 m	Myurella	affinis (JE Gray 1834)	X	X	X	III	E
30511	Panglao 04	9°35.7′N, 123°44.4′E, 0–2 m	Terenolla	pygmaea (Hinds 1844)	X	X	X	III	E
30510	Panglao 04	9°35.7′N, 123°44.4′E, 0–2 m	Myurella	columellaris (Hinds 1844)	X	X		III	E
30459	Panglao 04	9°35.7′N, 123°44.4′E, 0–2 m	Myurella	kilburni (RD Burch 1965)	X	X	X		E
30460	Panglao 04	9°35.7′N, 123°44.4′E, 0–2 m	Myurella	affinis (JE Gray 1834)	X	X	X	III	E
30587	Panglao 04	9°37.4′N, 123°46.9′E, 3–20 m	Acus	areolatus (Link 1807)	X	X	X	IA	В
30472	Panglao 04	9°41.8′N, 123°51.1′E, 10 m	Cinguloterebra	fenestrata type I		X	X	IIB	C
30534	Panglao 04	9°42.1′N, 123°51.4′E, 3–4 m		n. sp. Terryn I	X	X	X		E
30465	Panglao 04	9°29.4′N, 123°56.0′E, 15–20 m	Myurella	flavofasciata (Pilsbry 1921)	X	X	X	III	E
30408	Panglao 04	9°29.4′N, 123°56.0′E, 15–20 m	Myurella	nebulosa (GB Sowerby I 1825)	X	X	X		E
30450	Panglao 04	9°43.3′N, 123°48.8′E, 123–135 m	Hastulopsis	sp.	X	X	X		E
30407	Panglao 04	9°32.8′N, 123°45.9′E, 0–2 m	Impages	hectica (Linnaeus 1758)	X	X	X	IIA	D
30451	Panglao 04	9°36.8′N, 123°52.2′E, intertidal	"Terebra" textilis-group	textilis type IV	X	X	X		E
30431	Panglao 04	9°36.8′N, 123°52.2′E, intertidal	Terebra	laevigata (JE Gray 1834)	X	X	X	IIB	C
30454	Panglao 04	9°36.4′N, 123°53.8′E, 60–62 m	Hastulopsis	sp.	X	X	X		E
30513	Panglao 04	9°36.4′N, 123°53.8′E, 60–62 m	cf. Myurella	sp.	X	X	X		E
30455	Panglao 04	9°36.4′N, 123°53.8′E, 60–62 m	Clathroterebra	poppei (Terryn 2003)	X	X	X		E
30483	Panglao 04	9°32.8′N, 123°42.1′E, 3–35 m	Terebra	subulata (Linnaeus 1767)	X	X	X	IIB	E
30481	Panglao 04	9°35.7′N, 123°44.4′E, 0–2 m	Myurella	affinis (JE Gray 1834)	X	X	X	III	E
30401	Panglao 05	9°39.2′N, 123°47.5′E, 255–268 m	Clathroterebra	fortunei type II	X	X	X		E
30582	Panglao 05	9°39.2′N, 123°47.5′E, 255–268 m	"Terebra" textilis-group	textilis (Hinds 1844)	X		X		E
30418	Panglao 05	9°39.2′N, 123°47.5′E, 255–268 m	Cinguloterebra	fenestrata type II	X	X	X	IIB	C
30410	Panglao 05	9°39.2′N, 123°47.5′E, 255–268 m	Cinguloterebra	fenestrata type I	X	X	X	IIB	C
30395	Panglao 05	9°37.5′N, 123°40.2′E, 606–631 m	"Terebra" bathyrhaphe-group	jungi (Lai 2001)	X	X	X	IIB	A
30390	Panglao 05	9°29.4′N, 123°44.4′E, 271–318 m	Cinguloterebra	cf. fenestrata (Hinds 1844)	X	X	X	IIB	C
15724	Panglao 05	9°27.4′N, 123°49.4′E, 273–356 m	Cinguloterebra	cf. fujitai (Kuroda and Habe 1952)	X	X	X		C
16731	Panglao 05	9°30.1′N, 123°41.6′E, 356–396 m	"Terebra"	sp.	X	X	X		E
16735	Panglao 05	9°36.2′N, 123°43.8′E, 382–434 m	Cinguloterebra	cf. fenestrata (Hinds 1844)	X	X	X	IIB	C
30584	Panglao 05	9°34.3′N, 123°37.8′E, 729–733 m	"Terebra" bathyrhaphe-group	jungi (Lai 2001)	X	X	X	IIB	A
30406	Panglao 05	6°24.1′S, 156°20.4′E, 1045–1207 m	Cinguloterebra	sp.	X				C
30479	Salomon 2	7°13.8′ S, 158° 29.4′ E, 286–423 m	"Terebra" textilis-group	textilis type VII	X	X	X		E
30528	Salomon 2	7°43.5′S, 158°29.7′E, 336–341 m	"Terebra" textilis-group	textilis type VII		X	X		E

Evolution of the Venom Apparatus in the Terebridae 19

Table 2 Continued

MNHN N°	Cruise	Coordinates, Depth	Genus	Species (Author)	COI	16S	12S	Miller Type	Clade
30424	Salomon 2	8°36.7′S, 157°21.0′E, 150–160 m	"Terebra" textilis-group	textilis type VI		X	X		E
30391	Salomon 2	7°59.3′S, 157°33.3′E, 260 m	Clathroterebra	fortunei type II	X	X	X		E
30501	Salomon 2	8°25.5′S, 159°26.4′E, 543–593 m	"Terebra" bathyrhaphe-group	jungi (Lai 2001)		X	X	IIB	A
30487	Salomon 2	8°39.5′S, 157°23.0′E, 214–243 m	Cinguloterebra	lima (Deshayes 1857)	X	X	X		C
30579	Salomon 2	8°39.5′S, 157°23.0′E, 214–243 m	"Terebra" textilis-group	trismacaria (Melvill 1917)	X	X	X		E
30492	Salomon 2	8°39.5′S, 157°23.0′E, 214–243 m	"Terebra" textilis-group	trismacaria (Melvill 1917)	X	X	X		E
30499	Salomon 2	8°39.5′S, 157°23.0′E, 214–243 m		Unidentified juvenile	X	X	X		E
30409	Santo 06	15°33.1′S, 167°17.8′E, 15–25 m	Terebra	tricolor (GB Sowerby I 1825)	X	X	X	IIB	C
30389	Santo 06	15°28.7′S, 167°15.2′E, 19 m	Acus	maculatus (Linnaeus 1758)	X	X	X	IB	В
30378	Santo 06	15°33.1′S, 167°12.2′E, 3–40 m	Myurella	nebulosa (GB Sowerby I 1825)		X	X		E
30629	Santo 06	15°33.1′S, 167°12.2′E, 3–40 m	Myurella	undulata (JE Gray 1834)	X	X	X	III	E
30628	Santo 06	15°33.1′S, 167°12.2′E, 3–40 m	Myurella	undulata (JE Gray 1834)	X	X	X	III	E
30626	Santo 06	15°31.7′S, 167°09.7′E, 18–21 m	"Terebra" textilis-group	textilis type V	X	X	X		E
30440	Santo 06	15°31.7′S, 167°09.7′E, 18–21 m	"Terebra" textilis-group	textilis type V	X	X	X		E
30456	Santo 06	15°31.7′S, 167°09.7′E, 18–21 m	"Terebra" textilis-group	textilis type V	X	X	X		E
30458	Santo 06	15°31.7′S, 167°09.7′E, 18–21 m	"Terebra" textilis-group	textilis type V	X	X	X		E
30634	Santo 06	15°31.7′S, 167°09.7′E, 18–21 m	"Terebra" textilis-group	textilis type V	X	X	X		E
30617	Santo 06	15°31.7′S, 167°09.7′E, 18–21 m	"Terebra" textilis-group	textilis type V	X	X	X		E
30618	Santo 06	15°31.7′S, 167°09.7′E, 18–21 m	"Terebra" textilis-group	textilis type V	X	X	X		E
30386	Santo 06	15°36.6′S, 167°10.1′E, 8–20 m	Terebra	subulata (Linnaeus 1767)	X	X	X	IIB	C
30619	Santo 06	15°33.1′S, 167°12.2′E, 3–40 m	Hastulopsis	conspersa (Hinds 1844)	X	X	X	III	E
30387	Santo 06	15°33.1′S, 167°12.2′E, 3–40 m	Terebra	guttata (Röding 1798)	X	X	X	IIB	C
30376	Santo 06	15°33.1′S, 167°12.2′E, 3–40 m	Terebra	guttata (Röding 1798)	X	X	X	IIB	C
30620	Santo 06	15°31.3′S, 167°10.4′E, 3–18 m	Myurella	undulata (JE Gray 1834)	X	X	X	III	E
30494	Santo 06	15°34.4′S, 167°13.1′E, 9 m	Acus	crenulatus (Linnaeus 1758)	X	X	X	IA	В
30377	Santo 06	15°34.4′S, 167°13.1′E, 9 m	Acus	crenulatus (Linnaeus 1758)	X	X	X	IA	В
30623	Santo 06	15°36.9′S, 167° 10.5′E, 6–33 m	Hastulopsis	conspersa (Hinds 1844)	X	X	X	III	E
30480	Santo 06	15°36.9′S, 167° 10.5′E, 6–33 m	Hastulopsis	amoena (Deshayes 1859)	X	X	X		E
30463	Santo 06	15°36.9′S, 167° 10.5′E, 6–33 m	Hastulopsis	conspersa (Hinds 1844)	X	X	X	III	E
30478	Santo 06	15°36.9′S, 167° 10.5′E, 6–33 m	Hastulopsis	conspersa (Hinds 1844)	X	X	X	III	E
30388	Santo 06	15°36.9′S, 167° 10.5′E, 6–33 m	Hastulopsis	pertusa (Born 1778)	X	X	X	III	E
30392	Santo 06	15°36.9′S, 167° 10.5′E, 6–33 m	Hastulopsis	pertusa (Born 1778)	X	X	X	III	E
30373	Santo 06	15°36.9′S, 167° 10.5′E, 6–33 m	Acus	dimidiatus (Linnaeus 1758)	X	X	X	IA	В
30394	Santo 06	15°36.9′S, 167° 10.5′E, 6–33 m	Terebra	laevigata (JE Gray 1834)	X	X	X	IIB	C
30475	Santo 06	15°36.9′S, 167° 10.5′E, 6–33 m	Terebra	laevigata	X	X	X		E
30476	Santo 06	15°36.9′S, 167° 10.5′E, 6–33 m	"Terebra" textilis-group	textilis type III	X	X	X		E
30621	Santo 06	15°36.9′S, 167° 10.5′E, 6–33 m	"Terebra" textilis-group	textilis type IV	X	X	X		E
30616	Santo 06	15°36.9′S, 167° 10.5′E, 6–33 m	"Terebra" textilis-group	textilis type IV	X	X	X		E
30558	Santo 06	15°31′S, 167°09′E, intertidal	Strioterebrum	plumbeum (Quoy and Gaimard 1833)	X	X	X		E
30469	Santo 06	15°31′S, 167°09′E, intertidal	Strioterebrum	plumbeum (Quoy and Gaimard 1833)	X	X	X		E
30632	Santo 06	15°31′S, 167°09′E, intertidal	Terebra	laevigata (JE Gray 1834)	X	X	X	IIB	C
30614	Santo 06	15°31′S, 167°09′E, intertidal	Strioterebrum	cf. arabella (Thiele 1925)		X	X		E
30613	Santo 06	15°31′S, 167°09′E, intertidal	Terebra	laevigata (JE Gray 1834)	X	X	X	IIB	C
30567	Santo 06	15°34.7′S, 167°13.8′E, 14–25 m	Myurella	nebulosa (GB Sowerby I 1825)	X	X	X		E
30374	Santo 06	15°31.4′S, 167°09.7′E, intertidal	Terebra	punctatostriata (JE Gray 1834)		X	X		C
30612	Santo 06	15°29′S, 167°14.94′E, 2–4 m	Myurella	parkinsoni (Bratcher and Cernohorsky 1976)	X	X	X	III	E
30471	Santo 06	15°29′S, 167°14.94′E, 2–4 m	Myurella	parkinsoni (Bratcher and Cernohorsky 1976)	X	X	X	III	E
30611	Santo 06	15°35.2′S, 167°59.4′E, intertidal	Strioterebrum	plumbeum (Quoy and Gaimard 1833)	X	X	X		E

Table 2 Continued

MNHN N°	Cruise	Coordinates, Depth	Genus	Species (Author)	COI	16S	12S	Miller Type	Clade
30610	Santo 06	15°35.2′S, 167°59.4′E, intertidal	Strioterebrum	cf. Plumbeum (Quoy and Gaimard 1833)	X	x	X		E
30609	Santo 06	15°35.2′S, 167°59.4′E, intertidal	Strioterebrum	sp.	X	X	X		E
30608	Santo 06	15°35.2′S, 167°59.4′E, intertidal	Hastula	strigilata (Linnaeus 1758)	X	X		IIA	D
30607	Santo 06	15°35.2′S, 167°59.4′E, intertidal	Hastula	strigilata (Linnaeus 1758)		X	X	IIA	D
30420	Santo 06	15°35.2′S, 167°59.4′E, intertidal	Hastula	strigilata (Linnaeus 1758)	X	X	X	IIA	D
30416	Santo 06	15°35.2′S, 167°59.4′E, intertidal	Hastula	strigilata (Linnaeus 1758)		X	X	IIA	D
30414	Santo 06	15°26.6′S, 167°15.2′E, intertidal	Myurella	affinis (JE Gray 1834)	X	X	X	IIA	Е
30412	Santo 06	15°26.6′S, 167° 15.2′E, intertidal	Myurella	affinis (JE Gray 1834)	X	X	X	III	Е
30598	Santo 06	15°26.6′S, 167° 15.2′E, intertidal	Myurella	columellaris (Hinds 1844)	X	X	X	III	Е
30385	Santo 06	15°26.6′S, 167° 15.2′E, intertidal	"Terebra" textilis-group	textilis (Hinds 1844)	X	X	X		Е
30597	Santo 06	15°26.6′S, 167° 15.2′E, intertidal	Terebra	laevigata (JE Gray 1834)	X	X	X	IIB	C
30384	Santo 06	15°26.6′S, 167° 15.2′E, intertidal	Myurella	undulata (JE Gray 1834)		X	X	III	E
30383	Santo 06	15°26.6′S, 167° 15.2′E, intertidal	Terebra	argus (Hinds 1844)		X	X	IIB	C
30382	Santo 06	15°26.6′S, 167° 15.2′E, intertidal	Terebra	cingulifera (Lamarck. 1822)	X	X	X	IIB	C
30552	Santo 06	15°35.2′S, 167°59.4′E, intertidal	Cinguloterebra	anilis (Röding 1798)		X	X	IIB	C
30375	Santo 06	15°31.1′S, 167°10.5′E, 7 m	Terebra	babylonia (Lamarck 1822)	X	X	X	IIB	C
30380	Santo 06	15°31.1′S, 167°10.5′E, 7 m	Terebra	babylonia (Lamarck 1822)	X	X	X	IIB	C
30594	Santo 06	15°31.1′S, 167°10.5′E, 7 m	Myurella	affinis (JE Gray 1834)	X	X	X	III	Е
30550	Santo 06	15°26.6′S, 167° 15.2′E, intertidal	Hastula	solida (Deshayes 1857)		X	X	IIA	D
30549	Santo 06	15°26.6′S, 167° 15.2′E, intertidal	Hastula	solida (Deshayes 1857)		X	X	IIA	D
30417	Santo 06	15°26.6′S, 167° 15.2′E, intertidal	Hastula	solida (Deshayes 1857)	X	X	X	IIA	D
30437	Santo 06	15°26.6′S, 167° 15.2′E, intertidal	Hastula	albula (Menke 1843)	X	X		IIA	D
30438	Santo 06	15°26.6′S, 167° 15.2′E, intertidal	Hastula	albula (Menke 1843)		X	X	IIA	D
30571	Santo 06	15°35.9′S, 167°01.3.1′E, 83–90 m	Clathroterebra	fortunei type I?	X	X	X		E
30498	Santo 06	15°27.6′S, 167°14.3′E, 6–35 m	Myurella	nebulosa (GB Sowerby I 1825)	X	X	X		E
30453	Santo 06	15°29.6′S, 167°14.9′E, 2–5 m	Myurella	paucistriata (EA Smith 1873)	X	X	X	III	E
30546	Santo 06	15°35.5′S, 167°02.7′E, 86–118 m	Clathroterebra	poppei (Terryn 2003)	X	X	X		Е
30544	Santo 06	15°28.6′S, 167°15.1′E, 3–31 m	Cinguloterebra	jenningsi (RD Burch 1965)	X	X	X	IIB	C
30603	Santo 06	15°43.4′S, 167°15.0′E, 6 m	Terebra	laevigata (JE Gray 1834)	X	X	X	IIB	C
30541	Santo 06	15°36.8S, 167°08.5′E, 1–42 m	Myurella	affinis (JE Gray 1834)	X	X	X	III	E
30379	Santo 06	15°32.5′S, 167°10.5′E, 5–10 m	Acus	dimidiatus (Linnaeus 1758)	X	X	X	IA	В
30372	Santo 06	15°32.5′S, 167°10.5′E, 5–10 m	Acus	dimidiatus (Linnaeus 1758)	X	X	X	IA	В
30493	Santo 06	15°38.5′S, 167°15.1′E, 13 m	Terebra	tricolor (GB Sowerby I 1825)	X	X	X	IIB	C
30485	Santo 06	15°32.5′S, 167°10.5′E, 5–10 m	Cinguloterebra	lima (Deshayes 1857)	X	X	X		C
30461	Santo 06	15°42.7′S, 167°09.6′E, 2–3 m	Myurella	kilburni (RD Burch 1965)	X	X	X		Е
30601	Santo 06	15°40.7′S, 167°0.5′E, 517–614 m	"Terebra " textilis-group	textilis type V	X	X	X		E
30570	Santo 06	15°38.1′S, 167°05.9′E, intertidal	Myurella	undulata (JE Gray 1834)	X	X	X	III	Е
30428	Santo 06	15°38.1′S, 167°05.9′E, intertidal	Acus	dimidiatus (Linnaeus 1758)	X	X	X	IA	В
30547	Santo 06	15°31.3′S, 167°09.91′E, 1–6 m	"Terebra " textilis-group	textilis type III	X	X	X		Е
30545	Santo 06	15°31.3′S, 167°09.91′E, 1–6 m	"Terebra " textilis-group	textilis type III	X	X	X		Е
30426	Santo 06	15°35.4′S, 166°58.7′E, 3–8 m	Impages	hectica (Linnaeus 1758)	X	X	X	IIA	D
30543	Santo 06	15°35.4′S, 166°58.7′E, 3–8 m	"Terebra " textilis-group	textilis (Hinds 1844)	X	X	X		Е
30457	Santo 06	15°22.6'S, 167° 11.6'E, intertidal	Hastula	cf. rufopunctata (EA Smith 1877)	X	X	X	IIA	D
30542	Santo 06	15°22.6'S, 167° 11.6'E, intertidal	Hastula	penicillata (Hinds 1844)	X	X	X	IIA	D
30540	Santo 06	15°22.6'S, 167° 11.6'E, intertidal	Hastula	penicillata (Hinds 1844)	X	X	X	IIA	D
30490	Santo 06	15°22.6'S, 167° 11.6'E, intertidal	Acus	chloratus (Lamarck 1822)	X	X	X	IB	В
30425	Santo 06	15°35.4′S, 166°58.7′E, 3–8 m	Strioterebrum	cf. arabella (Thiele 1925)	X	X	X		E
30473	Santo 06	15°35.4′S, 166°58.7′E, 3–8 m	Strioterebrum	cf. arabella (Thiele 1925)	X	X	X		E

MNHN N°	Cruise	Coordinates, Depth	Genus	Species (Author)	COI	168	12S	COI 16S 12S Miller Type	Clade
30474	Santo 06	15°35.4′S, 166°58.7′E, 3–8 m	"Terebra" textilis-group	textilis type III	×	X	X		Ε
17938	Santo 06	15°35.4′S, 166°58.7′E, 3–8 m	"Terebra" textilis-group	textilis (Hinds 1844)	×	×	×		田
30381	Santo 06	15°35.4′S, 166°59.7′E, 3–37 m	Acus	dimidiatus (Linnaeus 1758)	×	×	×	IA	В
30630	Santo 06	15°35.7'S, 166°59.3'E, 12 m	Hastula	albula (Menke 1843)	×	×	×	IIA	О
30433	Santo 06	15°33.4′S, 167°12.4′E, 2–6 m	"Terebra" textilis-group	textilis type V	×	×	×		田
30434	Santo 06	15°33.4′S, 167°12.4′E, 2–6 m	"Terebra" textilis-group	textilis type V	×	×	×		田
30624	Santo 06	15°33.4′S, 167°12.4′E, 2–6 m	Hastulopsis	conspersa (Hinds 1844)	×	×	×	Ш	田
30635	Santo 06	15°33.4′S, 167°12.4′E, 2–6 m	Myurella	parkinsoni (Bratcher and Cernohorsky 1976)	×	×	×	Ш	田
30423	Santo 06	15°33′S, 167°16.7′E, 92 m	"Terebra" textilis-group	textilis type IV	×	×	×		凶
30403	Suva 4	15°26.4′S, 178°02.4′E, 50–51 m	Cinguloterebra	cumingii (Deshayes 1857)			×		C
30409	Suva 4	15°26.4′S, 178°02.4′E, 50–51 m	Cinguloterebra	cumingii (Deshayes 1857)		×	×		C
									l

sequencer. All genes were sequenced in both directions for increased accuracy of each sequence. Sequences deposited in GenBank (accession numbers: EU127881, EU015734, and EU685339–EU685783). COI sequences were also deposited in Barcode of Life Data Systems (http://www.barcodinglife.org/views/taxbrowser. php?taxid=897).

Phylogenetic Analyses

COI sequences were manually aligned, and 12S and 16S were automatically aligned using ClustalW multiple alignment implemented using default parameters in Bio-Edit version 7.0.5.3 (Hall 1999). Visual inspection was used to search the automatic alignments for any obvious anomalies. Hypervariable regions of 12S and 16S genes were excluded from further analyses due to ambiguities in the alignments. Nucleotide substitution models were selected for each gene separately using the program Modeltest (Posada and Crandall 2001) in conjunction with PAUP 4.0b10 (Swofford 2002). The best model of evolution and parameters estimated for each gene are Tamura-Nei (TrN) + I (0.537) + G (0.885) for COI gene, transversion model (TVM) + I(0.310) + G(0.581) for 12S gene, and general time reversible (GTR) + I (0.402) + G (0.483) for 16S gene. Phylogenetic analyses were based on reconstructions using maximum likelihood (ML) and Bayesian analysis (BA). The ML heuristic search was conducted with 100 replicates with tree_bissection and reconnection (TBR) branch swapping using PhyML 2.4.4 (Guindon and Gascuel 2003), and robustness of the nodes was assessed using nonparametric bootstrapping (Felsenstein 2004) with 1,000 bootstrap replicates for ML analysis.

BA consisted of six Markov chains (10,000,000 generations each with a sampling frequency of one tree each hundred generations) run in two parallel analyses using MrBayes (Huelsenbeck et al. 2001). The number of swaps that are tried each time the chain stops for swapping was four, and the chain temperature was set at 0.08. When the log-likelihood scores were found to stabilize, a consensus tree was calculated after omitting the first 0.25% of trees as burn-in.

For the combined analyses of the three genes, the same parameters were used for the ML analysis. For the BA, one different model was applied for each gene, each with six substitution categories. For the COI gene, as saturation was found on the third base of the codon, different models were applied for the two partitions (bases 1 and 2 vs. base 3). Finally, we have four unlinked partitions (COI bases 1 and 2, COI base 3, 12S, and 16S).

Venom Apparatus Mapping

Data from cited literature and personal communications from A. Sysoev and J. Taylor helped to determine the presence or absence of a venom apparatus in the species used for the phylogenetic analysis. The absence or presence of the venom apparatus was then mapped on the tree using Mesquite V. 2.01 (Maddison W and Maddison DR 2007), using the option "tracing character history." The parsimony ancestral reconstruction method was used.

Results

Phylogenetic Analyses

One hundred and fifty six samples of terebrids were used to reconstruct the molecular phylogeny of the Terebridae. For the COI gene, 658 bp were sequenced. After alignments, we obtained a fragment of 534 and 455 bp for 12S and 16S genes, respectively. Trees obtained independently with COI, 12S, and 16S genes were only partly resolved, but no contradictions were found (results not showed). Consequently, 12S, 16S, and COI mitochondrial genes, including 131 taxa in the ingroup, were used to produce a combined data set for phylogenetic analyses (fig. 2).

Phylogenetic analyses strongly indicate that the Terebridae is monophyletic (posterior probabilities [PP] = 1, bootstraps [B] = 100) (fig. 2). Terebra s.l. "Terebra" jungi appears to be the sister group to all other terebrids (PP = 1, B = 92). Apart from "T." jungi (Clade A), there are four major clades within the tree, designated in figure 2 as Clades B–E. Clade B is comprised primarily of the genus Acus plus one species currently placed in Terebra, Terebra areolata, which because of its placement in our tree, we tentatively define as Acus areolatus (PP = 1, B = 100). Clade C includes Cinguloterebra and additional species currently placed in Terebra, including the type species of Terebra, Terebra subulata (PP = 1, B = 76). Clade D includes species currently placed in the genera Hastula and Impages (PP = 1, B = 100).

Clade E is the largest clade and includes the genera Myurella, Clathroterebra, Terenolla, Hastulopsis. Strioterebrum, and the "Terebra" textilis-group (Terryn 2007). Myurella itself is polyphyletic, with several species placed as sister groups of other genera. For example, Myurella affinis, the type species, is the sister group of Terenolla, Hastulopsis, some Clathroterebra species, and two other Myurella species (PP = 1, B = 94). Clathroterebra is also polyphyletic, with the two representative species used in our analyses, Clathroterebra poppei and Clathroterebra fortunei, type species of Clathroterebra, appearing in separate well-supported clades (PP = 1, B = 100) for both. The distinctiveness of the monotypic genus *Terenolla* appears to be confirmed in our analysis. Hastulopsis and Strioterebrum are paraphyletic. The "Terebra" textilis-group is dispersed within Clade E and constitutes a group that includes a large amount of undescribed diversity, both at the genus and species levels, hence the various types (textilis III, IV, V, and VII) listed in the tree.

Evolution of Venom Apparatus

Miller's type classifications are indicated in the tree in figure 2 and table 2. "Terebra" jungi has a venom apparatus and is identified as a Type IIB feeder (Sysoev A, personal communication). Clade B corresponds to Miller's Type I feeders, which were further subdivided between Type IA and Type IB feeders. Type IA feeders are Acus crenulatus and Acus dimidiatus (Miller 1970, 1975). Acus areolatus, not hitherto recognized as a species of Acus, is however similar to the other species in Clade B in not having a venom apparatus and was identified as a Type IA feeder by Miller

(1975). The Type IB feeders in Clade B are Acus felinus, Acus maculatus, and Acus chloratus. The taxa that make up Clade C are all Type IIB feeders: Terebra guttata, T. subulata, Terebra babylonia, Terebra tricolor, Cinguloterebra fenestrata, Cinguloterebra anilis, and Cinguloterebra jenningsi (Taylor J, personal communication); (Miller 1970; Taylor 1990). Clade D made up exclusively of members of the genera Hastula (Hastula strigilata, Hastula penicillata, Hastula solida, Hastula albula, Hastula cf. rufopunctata) and Impages (Impages hectica) are Type IIA feeders (Miller 1979; Taylor 1990). Type III feeders are represented in the phylogenetic tree by Clade E, which includes species of Myurella (Myurella columellaris, Myurella undulata, Myurella paucistriata, M. affinis, Myurella flavofasciata, and Myurella parkinsoni), Hastulopsis (Hastulopsis conspersa and Hastulopsis pertusa), and Terenolla (Terenolla pygmaea) (Taylor J, personal communication) (Miller 1970; Taylor 1990).

In order to efficiently characterize toxins from various terebrid species, it is essential to first identify those species that have a venom apparatus. Shown in figure 3 is the mapping of the presence or absence of a venom apparatus in the terebrid species used to construct the molecular phylogeny in figure 2. The map clearly indicates that terebrids have lost the venom apparatus twice during their evolution, see Clades B and E. "T." jungi, and the members of Clades C and D, use the typical toxoglossate venom apparatus to hunt prey.

Discussion

The first molecular phylogeny of the Terebridae, presented in this report, significantly updates current hypotheses about the evolution of this group and will facilitate the efforts to characterize terebrid toxins. Although covering approximately 20% of the species-level diversity of the family, representing 12 of the 15 currently accepted genera, our data set does not include representatives of *Duplicaria* or *Pervicacia*, the latter made the type of a distinct family Pervicaciidae by Rudman (1969). However, our results highlight several problematic propositions about the Terebridae, while confirming the established hypotheses: 1) Terebridae family is monophyletic. 2) Shell shape was intuitively an appropriate character to group all terebrids in one taxon. Another substantiated hypothesis is that the monotypic genus Terenolla belongs in the Terebridae and is not a columbellid as suggested earlier (Bratcher and Cernohorsky 1987). A distinctive discovery revealed in figure 2 is that "T." jungi is a sister group to all other terebrid species. This is an original finding unappreciated until the present molecular analysis.

This phylogeny implies that there is little congruence between the former genus-level classification based on shell morphology and clades recognized by molecular characters. The molecular phylogeny indicates that most of the genera proposed in the family Terebridae will have to be revised. This is evidenced by, for example, species earlier assigned to *Terebra* now appearing throughout the tree in Clades B ("*T.*" areolata, here classified in *Acus*), C (*T. subulata*, type species of *Terebra*, and several others), and E ("*Terebra*" textilis-group). In addition, "*T.*" jungi will require the

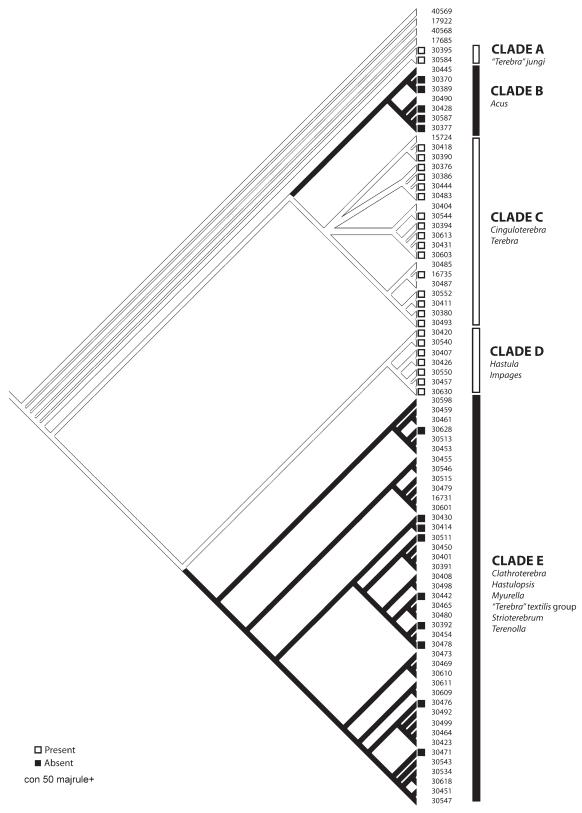


Fig. 3.—Possession of venom apparatus mapped onto Terebrid phylogeny. The presence or absence of a venom apparatus was mapped onto the molecular phylogeny of the Terebridae shown in figure 1. Terebrid species with a venom apparatus are indicated by a white box (\square), whereas terebrid species without a venom apparatus are indicated by a black box (). The map indicates that terebrids have independently lost the venom apparatus twice during their evolution.





Clade C

Fig. 4.—Terebrids with venom apparatus: representative shell images of the terebrid species in Clades C and D that have a venom apparatus. The species from left to right are Clade C: Terebra subulata, Terebra guttata, Cinguloterebra jenningsi, Cinguloterebra anilis, Terebra babylonia, Terebra laevigata and Clade D: Hastula strigilata, Hastula solida, and Hastula hectica.

establishment of a new genus (Terryn Y, Holford M, unpublished data). Despite this nonmonophyly of shell-defined genera, but based on the inclusion of their type species (except for Acus) in the analysis, we tentatively ascribe existing generic names to the four major clades: Clade B is thus Acus H. & A. Adams 1853, Clade C is *Terebra* Bruguière 1789, Clade D is *Hastula* H. & A. Adams 1853, and Clade E is Myurella Hinds 1844.

Evolutionary trends in the Toxoglossa have been reconstructed primarily through studies involving radular formation and anatomy of the digestive system (Mills 1979; Shimek and Kohn 1981; Kantor and Sysoev 1989; Taylor 1990; Simone 1999; Kantor and Taylor 2000). The molecular phylogeny presented here paints a plausible picture of how terebrids evolved such a diversity of feeding strategies. Our findings suggest that all terebrids appear to be derived from a common ancestor with a venom apparatus of the Miller Type IIB (fig. 3). Furthermore, mapping of the venom apparatus indicates that two lineages of terebrids independently lost their ability to hunt prey using toxins, Clades B and E.

There is a considerable correlation between our molecular phylogeny, Miller's anatomical groupings, and the ecological distribution of the terebrid species used in this study. Miller separated terebrids with a venom apparatus, Type II feeders, into two distinct groups, IIA and IIB. Type IIA and IIB terebrids differ in the shape of the buccal tube and shell morphology (Miller 1970). Type IIA terebrids have a long and slender buccal tube and small shiny shells, with 7–10 whorls and a flared aperture. Terebrids of Type IIB have a short, thick buccal tube, and the shells are large, long, and slender, with 15 or more whorls and a constricted aperture. Our phylogenetic analysis supports this separation. The species in Clade C, Terebra, have slender and multiwhorled shells, whereas those of Clade D, *Hastula*, are shiny with fewer whorls. The separation of the two clades is further supported by the ecological differences in their habitats. Terebra species of Clade C live buried in sandy or muddy subtidal flats, whereas Hastula species of Clade D live predominantly on surf beaches or in sand in reef pockets (Miller 1970, 1979). Similarly, terebrids that feed without the use of a venom apparatus, Types I and III feeders, are represented by two different clades in the molecular phylogeny, Clades B and E, respectively. Our analysis clarifies that the two clades without venom apparatus, Clades B and E, are not sister groups.

The Terebridae phylogeny in figure 2 sets the stage for efficient characterization of terebrid toxins and identification of the gene superfamilies that encode their toxins using the biodiversity first, exogenomic strategy recently applied to cone snails (Olivera 2006; Olivera and Teichert 2007). The exogenomic strategy was used to characterize cone snail toxins that target nicotinic receptors. In this strategy, phylogeny and molecular biology techniques are used to identify "exogenes," genes of the toxins expressed in the venom duct. Exogenes rapidly evolve to respond to cues in their biotic environment and are thus a powerful marker for differentiating ecological or evolutionarily distinct organisms.

Clades C and D are the two major terebrid groups most suitable to investigate toxins for biochemical characterization (fig. 4). Furthermore, as Clades C and D are not sister clades, they may produce divergent toxins that could result in varied functional activity upon further characterization.

Initial reports on the toxins from terebrids (Imperial et al. 2003, 2007) indicate that they have precursor structures organized similarly to those of conotoxin gene superfamilies, with a highly conserved signal sequence, followed by a propertide region, then the mature toxin region. As has been demonstrated with cone snails, the conserved signal sequence and propeptide region in terebrids are an exploitable feature that can be used to facilitate their characterization using molecular biology techniques, for example, as the basis for designing PCR primers. Preliminary results from the toxins characterized from T. subulata (Imperial et al. 2003) and Hastula hectica (Imperial et al. 2007) indicate that terebrid toxins are divergent from Conus toxins in several ways: 1) Terebrid toxins are not posttranslationally modified; facilitating the ability to chemically synthesize these peptides for testing of their function. 2) There is no homology between the gene superfamilies found in Conus and terebrid venom. These findings would indicate investigation of terebrid venom may reveal toxins with novel pharmacological specificity not found in Conus venom.

The biochemical and genetic characterization of terebrid toxins, while identifying novel compounds useful for investigating cell communication in the nervous system, will also provide additional characters to further clarify the phylogeny and evolutionary biology of these organisms. For toxoglossate gastropods, the dual analysis of molecular phylogeny and venom function is an instructive combination for unraveling the bigger question of evolutionary diversification. This work is a first attempt to address these issues for the Terebridae.

Acknowledgments

The authors thank Yuri Kantor for processing many of the terebrid specimens in the field, Virginie Heros and Philippe Maestrati for assistance throughout from the field to curating samples at MNHN, the staff of MNHN's "Service de Systématique Moléculaire" for technical facilities, and John Taylor and Alexander Sysoev for providing unpublished anatomical information on terebrid taxa used in this study. The Panglao 2004 and Santo 2006 expeditions, which were the source of many specimens, were supported among others by the Total Foundation. Joint funding from National Science Foundation Chemistry Division and Office of International Science and Engineering postdoctoral fellowship (0610202) for M.H. also supported this work.

Literature Cited

- Bouchet P. 1990. Turrid genera and mode of development: the use and abuse of protoconch morphology. Malacologia. 32:69-77.
- Bouchet P, Rocroi JP. 2005. Classification and nomenclature of gastropod families. Malacologia. 47(1-2):1-397.
- Bratcher T, Cernohorsky WO. 1987. Living terebras of the world. New York: American Malacologists, Inc.
- Darlu P, Tassy P. 1993. La reconstruction phylogénétique. Concepts et méthodes. Paris: Masson.
- Felsenstein J. 2004. Inferring phylogenies. Sunderland (MA): Sinauer Associates.
- Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R. 1994. DNA primers for amplification of mitochondrial Cytochrome C oxidase subunit I from diverse metazoan invertebrates. Mol Mar Biol Biotechnol. 3:294-299.
- Guindon S, Gascuel O. 2003. A simple, fast, and accurate algorithm to estimate large phylogenies by maximum likelihood. Syst Biol. 52:696-704.
- Hall T. 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/ NT. Nucleic Acids Symp Ser. 41:95-98.
- Herbert P, Cywinska A, Ball SL, deWaard JR. 2003. Biological identifications through DNA barcodes. Proc R Soc Lond B Biol Sci. 270:313-321.
- Huelsenbeck JP, Ronquist F, Hall B. 2001. MRBAYES: Bayesian inference of phylogeny. Bioinformatics. 17:753-755.
- Imperial J, Kantor Y, Watkins M, et al. (11 co-authors). 2007. Venomous auger snail Hastula (Impages) hectica (Linnaeus, 1758): molecular phylogeny, foregut anatomy, and comparative toxinology. J Exp Zool. 308B:1-13.
- Imperial JS, Watkins M, Chen P, Hillyard DR, Cruz LJ, Olivera BM. 2003. The augertoxins: biochemical characterization of venom components from the toxoglossate gastropod Terebra subulata. Toxicon. 41:391–398.
- Kantor Y, Sysoev AV. 1989. The morphology of toxoglossan gastropods lacking a radula, with a description of new species and genus of Turridae. J Molluscan Stud. 55:537-549.
- Kantor Y, Taylor JD. 2000. Formation of marginal radular teeth in Conoidea (Neogastropoda) and the evolution of the hypodermic envenomation mechanism. J Zool (Lond). 252:251-262.

- Maddison W, Maddison DR. 2007. Mesquite: a modular system for evolutionary analysis version 2.01.
- Miller BA. 1970. Studies on the biology of Indo-Pacific Terebra [PhD dissertation]. [Durham (NH)]: University of New Hampshire: 1970.
- Miller BA. 1971. Feeding mechanisms of the family Terebridae. Annu Rep Am Malacol Union. 1970:72-74.
- Miller BA. 1975. The biology of *Terebra gouldi* Deshayes, 1859, and a discussion of life history similarities among other terebrids of similar proboscis type. Pac Sci. 29(3):227-241.
- Miller BA. 1979. The biology of Hastula inconstans (Hinds, 1844) and a discussion of life history similarities among other Hastulas of similar proboscis type. Pac Sci. 33(3):289–306.
- Mills P. 1979. Radular tooth structure of three species of Terebridae (Mollusca: Toxoglossa). Veliger. 19:259–265.
- Olivera BM. 2002. Conus venom peptides: reflections from the biology of clades and species. Annu Rev Ecol Syst. 33:25-42.
- Olivera BM. 2006. Conus peptides: biodiversity-based discovery and exogenomics. J Biol Chem. 281:31173-31177.
- Olivera BM, Teichert RW. 2007. Diversity of the neurotoxic Conus peptides: a model for concerted pharmacological discovery. Mol Interv. 7(5):253-262.
- Oyama K. 1961. On some new facts of the taxonomy of Terebridae. Venus (Jpn J Malac). 21(2):176-189.
- Palumbi S. 1996. Nucleic Acids II: the polymerase chain reaction. In: Hillis D, Moritz C, Mable BK, editors. Molecular systematics. Sunderland (MA): Sinauer Associates. p. 205-247.
- Posada D, Crandall KA. 2001. Selecting models of nucleotide substitution: an application to human immunodeficiency virus (HIV-1). Mol Biol Evol. 18:897-906.
- Puillandre N, Samadi S, Boisselier MC, Sysoev AV, Kantor YI, Cruaud C, Couloux A, Bouchet P. 2008. Starting to unravel the toxoglossan knot: molecular phylogeny of the turrids (Neogastropoda: Conoidea). Mol Phylogenet 43(3):1122-1134.
- Rudman WB. 1969. Observations on Pervicacia tristis (Deshayes, 1859) and a comparison with other toxoglossan gastropods. Veliger. 12(1):53-64.
- Shimek RL, Kohn AJ. 1981. Functional morphology and evolution of the toxoglossan radula. Malacologia. 20:423-438.
- Simon C, Franke A, Martin A. 1991. The polymerase chain reaction: DNA extraction and amplification. In: Hewitt G, Johnson AWB, Young JPW, editors. Molecular techniques in taxonomy. New York: Springer-Verlag. H57. p. 329-355.
- Simone L. 1999. Comparative morphology and systematics of Brazilian Terebridae (Mollusca, Gastropoda, Conoidea), with descriptions of three new species. Zoosystema. 21(2):199-248.
- Swofford D. 2002. PAUP*: phylogenetic analysis using parsimony (*and other methods). Sunderland (MA): Sinauer Associates.
- Taylor JD. 1990. The anatomy of the foregut and relationships in the Terebridae. Malacologia. 32:19-34.
- Taylor JD, Kantor YI, Sysoev AV. 1993. Foregut anatomy, feeding mechanisms, relationships and classification of the Conoidea (= Toxoglossa) (Gastropoda). Bull Nat Hist Mus Lond (Zool). 59:125-170.
- Terryn Y. 2007. A collectors guide to recent Terebridae Neogastropoda). (Mollusca: Hackenheim (Germany): Conchbooks & NaturalArt.

Douglas Crawford, Associate Editor

Accepted September 10, 2008